

REPORTS

References and Notes

1. T. Pawson, J. D. Scott, *Science* **278**, 2075 (1997).
2. M. Sheng, *Proc. Natl. Acad. Sci. U.S.A.* **98**, 7058 (2001).
3. H. C. Kornau, L. T. Schenker, M. B. Kennedy, P. H. Seeburg, *Science* **269**, 1737 (1995).
4. J. E. Brenman et al., *Cell* **84**, 757 (1996).
5. V. L. Dawson, T. M. Dawson, E. D. London, D. S. Bredt, S. H. Snyder, *Proc. Natl. Acad. Sci. U.S.A.* **88**, 6368 (1991).
6. M. Migaud et al., *Nature* **396**, 433 (1998).
7. R. Sattler et al., *Science* **284**, 1845 (1999).
8. R. P. Simon, J. H. Swan, B. S. Meldrum, *Science* **226**, 850 (1984).
9. A. S. Fix et al., *Exp. Neurol.* **123**, 204 (1993).
10. S. M. Davis et al., *Stroke* **31**, 347 (2000).
11. G. F. Morris et al., *J. Neurosurg.* **91**, 737 (1999).
12. S. R. Schwarze, A. Ho, A. Vocero-Akbani, S. F. Dowdy, *Science* **285**, 1569 (1999).
13. See supporting data on Science Online.
14. D. A. Mann, A. D. Frankel, *EMBO J.* **10**, 1733 (1991).
15. Total EPSCs were recorded under conditions in which they are mediated by AMPARs (holding membrane potential of -60 mV, 2 mM extracellular Mg^{2+}).
16. Ratios of peak $[Ca^{2+}]_i$ and rate of rise of $[Ca^{2+}]_i$ before and after Tat-NR2B9c application were 1.03 ± 0.03 and 0.99 ± 0.04 , respectively ($n = 11$ neurons).
17. R. Sattler, M. P. Charlton, M. Hafner, M. Tymianski, *J. Neurochem.* **71**, 2349 (1998).
18. R. Sattler, Z. Xiong, W. Y. Lu, J. F. MacDonald, M. Tymianski, *J. Neurosci.* **20**, 22 (2000).
19. S. R. Jaffrey, S. H. Snyder, *Annu. Rev. Cell Dev. Biol.* **11**, 417 (1995).
20. Tat-NR2B9c-dansyl fluorescence was visualized in all other brain areas examined (hippocampus, striatum).
21. Tat-NR2B9c-dansyl fluorescence was detectable in brain vessels and all brain regions examined in 250-g Sprague-Dawley rats 1 hour after intravenous injection (3 nmol/g).
22. E. Z. Longa, P. R. Weinstein, S. Carlson, R. Cummins, *Stroke* **20**, 84 (1989).
23. L. Belayev, O. F. Alonso, R. Busto, W. Zhao, M. D. Ginsberg, *Stroke* **27**, 1616 (1996).
24. J. B. Bederson et al., *Stroke* **17**, 472 (1986).
25. M. De Ryck, J. van Reempts, M. Borgers, A. Wauquier, P. A. Janssen, *Stroke* **20**, 1383 (1989).
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Supporting Online Material

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Cancer Regression and Autoimmunity in Patients After Clonal Repopulation with Antitumor Lymphocytes

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We report here the adoptive transfer, to patients with metastatic melanoma, of highly selected tumor-reactive T cells directed against overexpressed self-derived differentiation antigens after a nonmyeloablative conditioning regimen. This approach resulted in the persistent clonal repopulation of T cells in those cancer patients, with the transferred cells proliferating in vivo, displaying functional activity, and trafficking to tumor sites. This led to regression of the patients' metastatic melanoma as well as to the onset of autoimmune melanocyte destruction. This approach presents new possibilities for the treatment of patients with cancer as well as patients with human immunodeficiency virus-related acquired immunodeficiency syndrome and other infectious diseases.

Immunotherapy of patients with cancer requires the in vivo generation of large numbers of highly reactive antitumor lymphocytes that are not restrained by normal tolerance mechanisms and are capable of sustaining immunity against solid tumors. Immunization of melanoma patients with cancer antigens can increase the number of circulating CD8⁺ cytotoxic T lymphocyte precursor cells (pCTLs), but to date

this has not correlated with clinical tumor regression, suggesting a defect in function or activation of the pCTLs (1).

Adoptive cell transfer therapies provide the opportunity to overcome tolerogenic mechanisms by enabling the selection and activation of highly reactive T cell subpopulations and by manipulation of the host environment into which the T cells are introduced. However, prior clinical trials, including the transfer of highly active antitumor T cell clones, failed to demonstrate engraftment and persistence of the transferred cells (2–5). Lymphodepletion can have a marked effect on the efficacy of T cell transfer therapy in murine models (6–

9) and may depend on the destruction of regulatory cells, disruption of homeostatic T cell regulation, or abrogation of other normal tolerogenic mechanisms.

To determine whether prior lymphodepletion might improve the persistence and function of adoptively transferred cells, 13 HLA-A2⁺ patients with metastatic melanoma received immunodepleting chemotherapy with cyclophosphamide and fludarabine for 7 days before the adoptive transfer of highly selected tumor-reactive T cells and high-dose interleukin-2 (IL-2) therapy (10) (Table 1). These patients all had progressive disease refractory to standard therapies, including high-dose IL-2, and eight patients also had progressive disease despite aggressive chemotherapy. The patients received an average of 7.8×10^{10} cells (range, 2.3×10^{10} to 13.7×10^{10}) and an average of nine doses of IL-2 (range, 5 to 12 doses). The T cells used for treatment were derived from tumor-infiltrating lymphocytes (TILs) and were rapidly expanded in vitro (11). All cultures were highly reactive when stimulated with an HLA-A2⁺ melanoma or an autologous melanoma cell line (Table 1 and table S1).

Six of the 13 patients had objective clinical responses to treatment and four others demonstrated mixed responses, with significant shrinkage of one or more metastatic deposits (11). Objective tumor regression was seen in the lung, liver, lymph nodes, and intraperitoneal masses and at cutaneous and subcutaneous sites. Five patients, all with evidence of concomitant cancer regression, demonstrated signs of autoimmune melanocyte destruction, including four patients with vitiligo and one patient with anterior uveitis (Table 1). All patients recovered from treatment with absolute neutrophil counts greater than $500/\text{mm}^3$ by day 11 after T cell infusion but with slower recovery of CD4⁺ cells, as expected after fludarabine therapy (12).

To investigate the function and fate of the transferred T cell populations, T cell receptor (TCR) expression was examined using a pan-

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el of beta chain variable region (V β)-specific antibodies in the six patients for whom peripheral blood samples were available at 1 week and approximately 1 month after cell transfer (table S2). V β expression was highly skewed in five of the six administered TILs, and these same V β families were also over-represented in the peripheral blood of the patients at 1 week after cell transfer. Patient 9 and patient 10 exhibited clinical and laboratory findings after treatment that represent the achievement of a primary goal of adoptive immunotherapy. Within a few days of cessation of IL-2 therapy after TIL transfer, patients 9 and 10 demonstrated a pronounced lymphocytosis, with absolute lymphocyte counts (ALCs) reaching peak levels in peripheral blood of over 21,000 cells/mm³ on day 7 after cell infusion for patient 9, and over 16,000 cells/mm³ on day 8 after cell infusion for patient 10 (Fig. 1A). PBLs from patient 9 sampled at the peak of the lymphocytosis were 94% CD8⁺, of which 63% expressed V β 12. Even more pronounced skewing of the T cell repertoire was observed in

PBLs of patient 10 sampled at the peak of lymphocytosis, when 96% of the lymphocytes were CD8⁺, of which 97% expressed V β 7 (Fig. 1B). Extreme skewing of TCR V β expression in the administered TIL and post-treatment PBL samples was confirmed using reverse transcription polymerase chain reaction (RT-PCR) with PCR primers that were designed to amplify all V β gene families (11). Thus, at both the RNA and protein levels, individual TCR V β families constituted a majority of the lymphocytes from the peripheral blood of patients 9 and 10 1 week after TIL transfer.

A V β 12⁺ T cell clone from the administered TILs of patient 9 and a V β 7⁺ clone from the TILs of patient 10, obtained by limiting dilution cloning and examined by cytokine secretion assay, exhibited specificity for the MART-1:27-35 epitope and strong recognition of HLA-A2⁺ melanoma cells (Fig. 1C). MART-1/melan-A is a normal, nonmutated differentiation antigen expressed on melanomas and normal melanocytes (13). The nucleotide sequence of the clonotypi-

cally unique TCR beta chain V(D)J regions was determined from MART-1 reactive V β 12⁺ and V β 7⁺ T cell clones from the TILs of patients 9 and 10 (Fig. 1D). Although the clones from both patients recognized the MART-1 antigen, no primary TCR sequence similarity in the antigen-combining region was evident. Additional nucleotide sequence analysis of the V(D)J regions of six DNA clones each from the infused TILs and the post-treatment PBLs of patient 9 demonstrated that the V β 12⁺ cells consisted of a clonally expanded cell population expressing the same MART-1-reactive TCR sequence shown in Fig. 1D. Similar results were obtained for the V β 7⁺ cells from TILs and post-treatment PBLs of patient 10 (11). Thus, clonal MART-1-reactive T cell populations within the TILs infused into patients 9 and 10 repopulated the immune systems of these patients. Furthermore, these results suggested that the individual clones had undergone significant expansion in vivo. Patient 9 had more than 5.0×10^{10} circulating V β 12⁺ lymphocytes; the patient was initially infused

Table 1. Patient demographics, treatments received, and clinical outcomes.

Patient	Age/sex	Treatment*				Sites of evaluable metastases	Response duration (months)	Autoimmunity
		Cells infused† ($\times 10^{-10}$)	CD8/CD4 phenotype‡ (%)	Antigen specificity§	IL-2 (doses)			
1	18/M	2.3	11/39	Other	9	Lymph nodes (axillary, mesenteric, pelvic)	PR¶ (24+)	None
2	30/F	3.5	83/15	MART-1, gp100	8	Cutaneous, subcutaneous	PR (8)	Vitiligo
3	43/F	4.0	44/58	gp100	5	Brain, cutaneous, liver, lung	NR	None
4	57/F	3.4	56/52	gp100	9	Cutaneous, subcutaneous	PR (2)	None
5	53/M	3.0	16/85	Other	7	Brain, lung, lymph nodes	NR-mixed	None
6	37/F	9.2	65/35	Other	6	Lung, intraperitoneal, subcutaneous	PR (15+)	None
7	44/M	12.3	61/41	MART-1	7	Lymph nodes, subcutaneous	NR-mixed	Vitiligo
8	48/M	9.5	48/52	gp100	12	Subcutaneous	NR	None
9	57/M	9.6	84/13	MART-1	10	Cutaneous, subcutaneous	PR (10+)	Vitiligo
10	55/M	10.7	96/2	MART-1	12	Lymph nodes, cutaneous, subcutaneous	PR¶ (9+)	Uveitis
11	29/M	13.0	96/3	MART-1	12	Liver, pericardial, subcutaneous	NR-mixed	Vitiligo
12	37/F	13.7	72/24	MART-1	11	Liver, lung, gallbladder, lymph nodes	NR-mixed	None
13	41/F	7.7	92/8	MART-1	11	Subcutaneous	NR	None

*Each patient was treated with chemotherapy (27) starting 7 days before cell administration, consisting of 2 days of cyclophosphamide at 60 mg per kg of body weight, followed by 5 days of fludarabine at 25 mg/m². On the day after the final dose of fludarabine, when circulating lymphocyte and neutrophil counts had dropped to less than 20/mm³, each patient received an intravenous infusion of autologous lymphocytes over approximately 30 to 60 min. After cell infusion, patients received high-dose IL-2 therapy consisting of 720,000 IU/kg by bolus intravenous infusion every 8 hours to tolerance (10). Some patients with mixed or responding lesions received an additional course of cell transfer therapy. † T cell cultures for infusion were derived from TILs by minor modifications of established techniques (2, 5). Multiple cultures were started from each resected melanoma specimen and were screened independently by cytokine secretion assay for recognition of autologous tumor cells (if available) and HLA-A2⁺ tumor cell lines (11). TIL cultures that exhibited specific tumor cell recognition were expanded for treatment using one or two cycles of a rapid expansion protocol (28) with irradiated allogeneic feeder cells, OKT3 (anti-CD3) antibody, and 6000 IU per ml of IL-2. ‡Percent of lymphocyte gated cells from the infusion sample that stained with each antibody. Values do not add to 100% if a significant fraction of infused cells was double negative or double positive for CD4 and CD8 antigen expression. §Antigen specificity was determined by cytokine release assay (11). Other: recognition of autologous tumor cells but no HLA-A2-restricted epitope derived from MART-1, tyrosinase, tyrosinase related protein 1 (TRP1), TRP2, NY-ESO-1, gp100, MAGE1, or MAGE3. ¶NR, no response; PR, partial response. ¶¶Microscopic residual focus of disease resected; patient remains free of disease (11). Patient 1 had a 6-mm brain density that increased to 8 mm at 8 months. After localized stereotactic radiotherapy, the density disappeared.

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with approximately 1.2×10^{10} V β 12⁺ TILs. Patient 10 had at least 5.6×10^{10} circulating V β 7⁺ lymphocytes; the patient was initially infused with 9.5×10^{10} V β 7⁺ TILs. Even without accounting for additional cells within lymphoid tissues or infiltrating into solid tissues, the preponderance of only a single clone in the peripheral blood of these two patients during their lymphocytic episodes was striking and indicated that the *in vivo* expansion of highly avid, functional antitumor lymphocytes had been achieved.

Although the absolute lymphocyte counts

decayed to normal levels after 3 to 4 weeks in patients 9 and 10, the composition of the resulting lymphocyte pool remained highly skewed for over 4 months. As measured by antibody to V β 12 and by A2/MART-1 tetramer fluorescence-activated cell sorting (FACS) (11) analysis, the MART-1-reactive clone in patient 9 remained above 60% of the CD8⁺ lymphocytes for over 123 days (Fig. 1E). Patient 10 retained the MART-1-reactive V β 7⁺ T cell at a level of more than 75% of the CD8⁺ cells throughout the course of the study (over 159 days from the date of TIL transfer).

High levels of specific lysis of MART-1:27-35 peptide pulsed targets (14) and MART-1-expressing HLA-A2⁺ tumor cell lines (Fig. 2A) were observed in the infused TILs and in the post-infusion PBLs. Blood smears of PBLs from patients 9 and 10 demonstrated that the circulating lymphocytes exhibited an atypical activated blastic morphology (Fig. 2B), consistent with their direct *ex vivo* lytic function. After overnight activation in IL-2 of post-transfer PBLs from patients 9 and 10, specific cytokine secretion to MART-1 peptide and HLA-A2⁺ tumors was

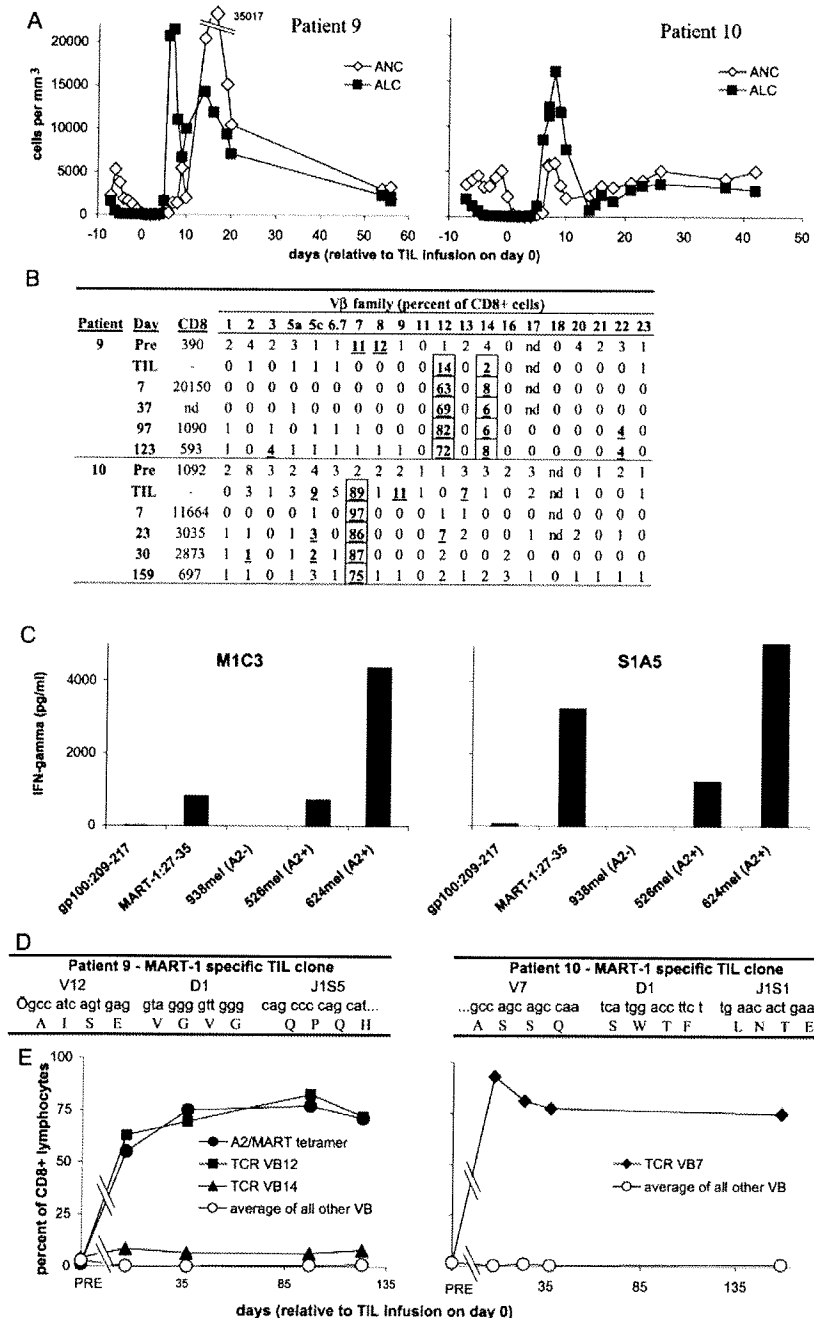


Fig. 1. (A) Patient 9 (left) and patient 10 (right) exhibited a profound lymphocytosis after treatment. ALCs (solid squares) and absolute neutrophil counts (ANCs) (open diamonds) are plotted over time. The upper limit of normal for ALCs is 4800 cells/mm³. **(B)** Individual V β s that were overrepresented in the infused cells (TILs) of patients 9 and 10 dominated the TCR repertoire of PBLs sampled after treatment (11). Values in the "CD8" column indicate the absolute number of CD8⁺ cells/mm³ of peripheral blood on the indicated day. PRE, pretreatment PBL; nd, not done. **(C)** T cell clones derived from the infused TILs by limiting dilution specifically recognized the MART-1:27-35 peptide and HLA-A2⁺ melanoma cell lines by cytokine secretion assay (11). M1C3 is a V β 12⁺ clone from patient 9 and S1A5 is a V β 7⁺ clone from patient 10. **(D)** The DNA and predicted protein sequences of complementarity-determining region 3 (CDR3) of the TCR β chains of a V β 12⁺ T cell clone from the infused TILs of patient 9 (left) or a V β 7⁺ T cell clone derived from the TILs of patient 10 (right) are shown. Although both clones recognize the MART-1 antigen, no CDR3 sequence similarity between the clones is evident. **(E)** MART-1-reactive clones persisted as the predominant lymphocyte population in the peripheral blood for more than 4 months. PBL samples from patient 9 (left) or patient 10 (right) at the indicated days after cell transfer were analyzed by FACS (11) using antibodies specific for the indicated TCR V β families (V β 7, solid diamonds; V β 12, solid squares; V β 14, solid triangles; average of other V β s, open circles) or HLA-A2/MART-1:26-35(27L) tetrameric complexes (A2/MART tetramer, solid circles). Lymphocytes from patient 10 reacted only with the native MART-1:27-35 peptide, not the altered MART-1:26-35(27L) peptide (14).

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evident, including interferon- γ (IFN- γ) (Fig. 2C), granulocyte-macrophage colony-stimulating factor, and tumor necrosis factor- α (14). These results suggested that the persistent *in vivo* expanded cells observed in both patients could express lytic activity as well as antigen-specific proinflammatory cytokine secretion after activation *in situ*.

Pretreatment tumor biopsy samples from patients 9 and 10 contained minimal lymphocytic infiltration (Fig. 3). After treatment, the biopsied specimens contained large areas of necrotic tumor and areas of dense, diffuse lymphocytic infiltrates. Immunohistochemical staining revealed that the lymphocytic

infiltrates consisted predominantly of CD8 $^{+}$ cells (Fig. 3, A and B). The infiltrating T cells from patient 9 were predominantly V β 12 $^{+}$ but not V β 7 $^{+}$, whereas T cells infiltrating tumor tissue from patient 10 were predominantly V β 7 $^{+}$ but not V β 12 $^{+}$ (Fig. 3A). RT-PCR and nucleotide sequence analysis of RNA from biopsied tumor specimens from patient 9 obtained 20 days after cell transfer demonstrated that the predominant V β 12 $^{+}$ cells expressed the MART-1-reactive TCR sequence (11). Both major histocompatibility complex (MHC) class I and MHC class II antigens were highly expressed in tumor deposits after therapy but were expressed only

at low levels or not at all in tumors before TIL treatment (Fig. 3B), which was probably the result of IFN- γ secretion by the infiltrating lymphocytes (15). Thus, the proliferating, adoptively transferred antitumor cells were capable of trafficking to and infiltrating into tumor deposits.

Six patients, including patients 9 and 10, exhibited significant regression of metastatic melanoma, and five patients exhibited the onset of antimelanocyte autoimmunity (Table 1 and fig. S1). Patient 9 exhibited regression of more than 95% of his cutaneous and subcutaneous melanoma, ongoing at over 10 months, and developed vitiligo on his forearms. At four months after cell infusion, patient 9, who was Epstein-Barr virus (EBV) seronegative before treatment, developed an EBV-related lymphoproliferative disease that has been reported in EBV-seronegative patients receiving allogeneic transplants (16, 17), and he is undergoing treatment for this problem. One other patient (patient 5) developed an opportunistic infection during treatment, a respiratory syncytial virus that lasted for 1 week and then cleared. Patient 10 exhibited regression of more than 99% of his nodal, cutaneous, and subcutaneous melanoma deposits, ongoing at over 9 months. Fourteen days after cell infusion, during the active regression of melanoma, patient 10 developed bilateral acute anterior uveitis, characterized by a fibrinous pupillary membrane that has responded to steroid eye drops, and he remains healthy with normal vision (fig. S1).

This high frequency and extended persistence of individual T cell clones reactive with nonmutated self-antigens in humans has not previously been seen. Limited persistence of transferred cultured T cells reactive with viral antigens such as cytomegalovirus and human immunodeficiency virus has been reported but rarely at levels exceeding a few percent of circulating cells (17–21), although higher levels have been observed during the acute phase of infectious mononucleosis (22, 23).

The clinical and immunological findings from the patients reported here contrast sharply with our previous experience in investigating the adoptive transfer of cloned CD8 $^{+}$ T cells (including 15 patients who received cloned cells after nonmyeloablative conditioning), which did not result in any objective responses or any persistence of the transferred cells (5), and our experience with gene-modified TILs, which did not persist at high levels (24). One factor limiting *in vivo* persistence of CD8 $^{+}$ T cell clones as compared to the rapidly expanded TIL cultures reported here is the potential role of the transferred CD4 $^{+}$ (Table 1) cells, which have been shown in murine and human studies to play a role in the persistence of CD8 $^{+}$ cells (19, 25). Other factors may include an inherent proliferative impediment of clones expanded *in vitro*, the diversity of cell reactiv-

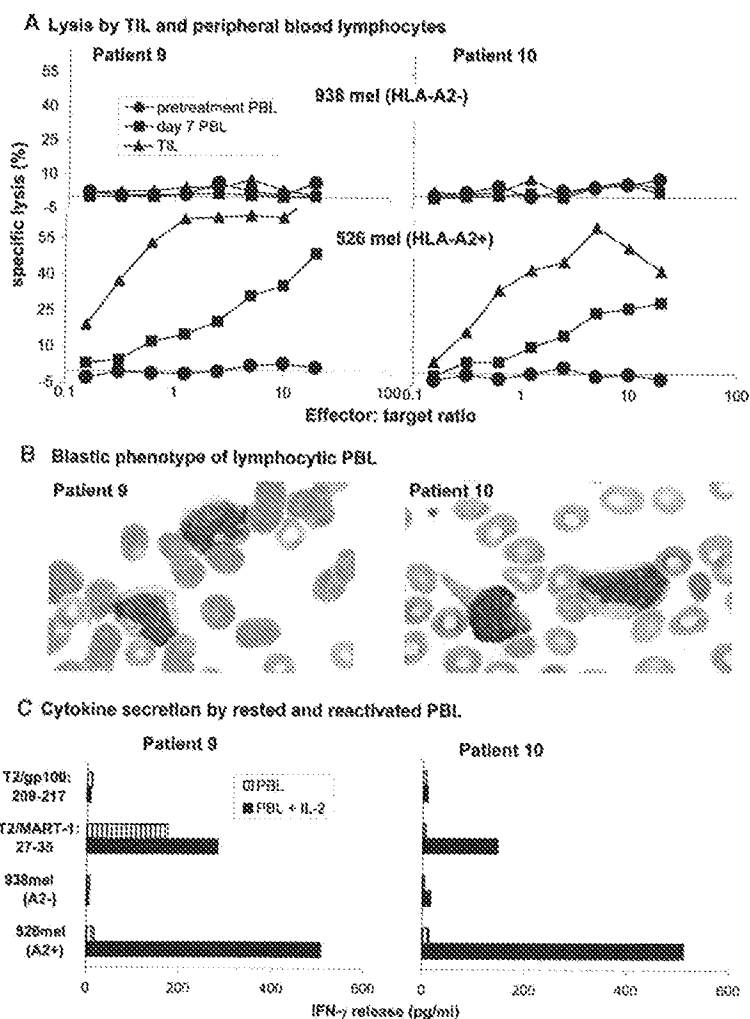


Fig. 2. (A) The infused TILs (triangles) and PBLs collected 7 days after cell transfer (squares), but not pretreatment PBLs (circles), from patient 9 (left) and patient 10 (right) specifically lysed the HLA-A2 $^{+}$ melanoma cell line 526 (lower graphs) but not the HLA-A2 $^{-}$ melanoma cell line 938 (upper graphs) (11). (B) Blood smears obtained from patient 9 (left) and patient 10 (right) during the lymphocytic episodes showed the highly activated phenotype of the lymphocytes, which show irregular and hyperchromatic nuclei, a high nuclear-to-cytoplasmic ratio, toxic granulation, and the presence of Dohle bodies. (C) PBLs collected 9 days (patient 9, left) or 8 days (patient 10, right) after cell transfer were cultured overnight in the absence (striped boxes) or presence (solid boxes) of 600 IU/ml of IL-2, washed extensively, and tested for cytokine release when stimulated with the indicated target cells.

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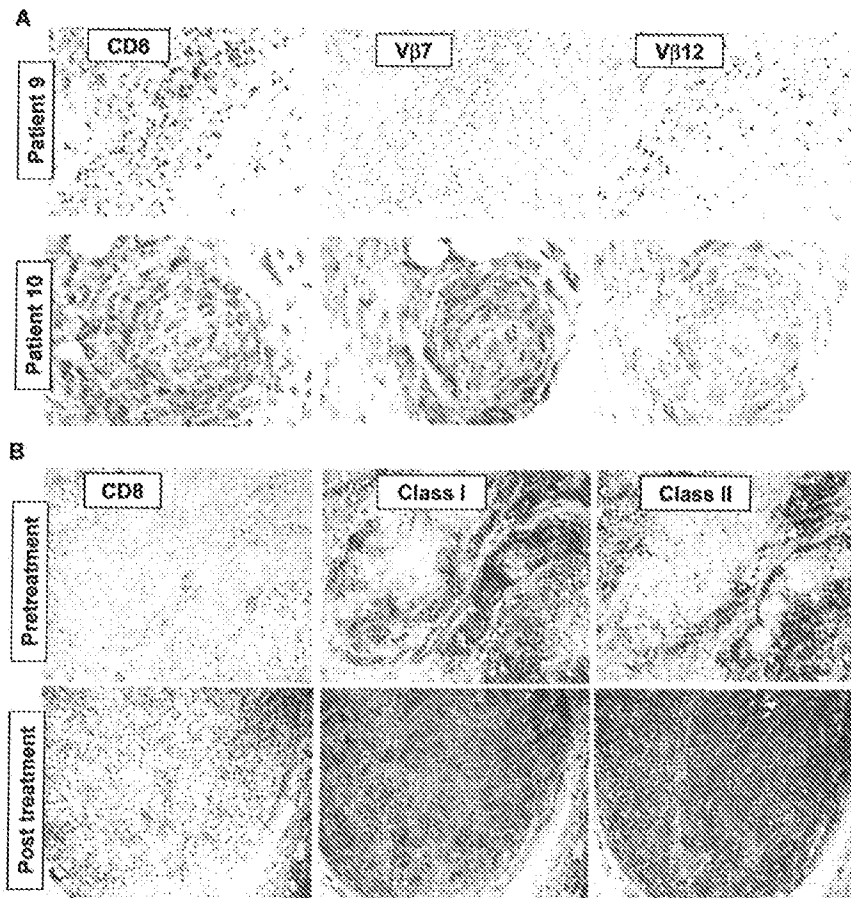


Fig. 3. (A) Antibodies specific for CD8 and TCR V β reveal tissue infiltration by a limited repertoire of T cells (17). Twenty days after cell transfer in patient 9, a resected tumor nodule contains diffusely infiltrating lymphocytes consisting of CD8⁺ V β 12⁺ T cells, but no V β 7⁺ T cells. Similarly, a residual tumor nodule biopsied 14 days after cell transfer from patient 10 exhibited few viable tumor cells but contained areas densely infiltrated by CD8⁺ V β 7⁺ T cells, with few V β 12⁺ cells. (B) A tumor lesion excised from patient 9 before nonmyeloablating chemotherapy ("pretreatment") exhibited scant CD8⁺ cells (left), strong stromal cell staining but weak staining of tumor cells with antibody to MHC class I (center), and sporadic cell staining with an antibody to MHC class II (probably tumor macrophages) but minimal staining of tumor cells (right). In contrast, a sample resected 57 days after cell transfer ("post treatment") exhibited a dense, diffuse CD8⁺ infiltrate and ubiquitous expression of both MHC class I and class II molecules in tumor cells.

ities transferred, or disordered trafficking after multiple anti-CD3-mediated expansions. The nonmyeloablative conditioning regimen may have eliminated regulatory cells or altered homeostatic mechanisms that limit lymphocyte numbers and thus played a role in enabling the in vivo proliferation of the transferred cells (26).

In summary, treatment with autologous T cell transfer and high-dose IL-2 therapy after nonmyeloablative lymphodepleting chemotherapy resulted in the rapid growth in vivo of clonal populations of T cells specific for the

MART-1 melanocyte differentiation antigen, and resulted in the destruction of metastatic tumors and autoimmune attack on normal tissues that expressed the MART-1 antigen. These cases demonstrate that adoptive transfer of tumor-reactive lymphocytes after nonmyeloablative conditioning can be an effective treatment for patients with metastatic cancer. Furthermore, this report establishes the principle that normally expressed "self-antigens" can be useful as targets for human tumor immunotherapy if the autoimmune consequences of such treatment are tolerable

or locally controllable, as may be the case for patients with a variety of common cancers such as those of the prostate, breast, ovary, or thyroid, the functions of which are not necessary for survival or can readily be replaced. A similar approach could be applied to other tumor histologies or to infectious diseases, such as acquired immunodeficiency virus, for which functionally active lymphocytes can be selected in vitro.

References and Notes

1. S. A. Rosenberg et al., *Nature Med.* **4**, 321 (1998).
2. S. A. Rosenberg et al., *J. Natl. Cancer Inst.* **86**, 1159 (1994).
3. C. Yee et al., *J. Exp. Med.* **192**, 1637 (2000).
4. M. E. Dudley et al., *J. Immunother.* **24**, 363 (2001).
5. M. E. Dudley et al., *J. Immunother.* **25**, 243 (2002).
6. J. R. Berenson, A. B. Einstein Jr., A. Fefer, *J. Immunol.* **115**, 234 (1975).
7. T. J. Eberlein, M. Rosenstein, S. A. Rosenberg, *J. Exp. Med.* **156**, 385 (1982).
8. R. J. North, *J. Exp. Med.* **155**, 1063 (1982).
9. S. A. Rosenberg, P. Spiess, R. Lafreniere, *Science* **233**, 1318 (1986).
10. S. A. Rosenberg, J. C. Yang, D. E. White, S. M. Steinberg, *Ann. Surg.* **228**, 307 (1998).
11. Materials and methods are available as supporting material on Science Online.
12. B. D. Cheson, *J. Clin. Oncol.* **13**, 2431 (1995).
13. Y. Kawakami et al., *Proc. Natl. Acad. Sci. U.S.A.* **91**, 3515 (1994).
14. M. E. Dudley et al., unpublished observations.
15. U. Boehm, T. Klamp, M. Groot, J. C. Howard, *Annu. Rev. Immunol.* **15**, 749 (1997).
16. R. J. O'Reilly et al., *Important Adv. Oncol.* **1996**, 149 (1996).
17. C. M. Rooney et al., *Blood* **92**, 1549 (1998).
18. S. R. Riddell et al., *Science* **257**, 238 (1992).
19. E. A. Walter et al., *N. Engl. J. Med.* **333**, 1038 (1995).
20. R. T. Mitsuyasu et al., *Blood* **96**, 785 (2000).
21. R. E. Walker et al., *Blood* **96**, 467 (2000).
22. M. F. Calian et al., *J. Clin. Invest.* **106**, 1251 (2000).
23. A. D. Hislop, N. E. Annels, N. H. Gudgeon, A. M. Leese, A. B. Rickinson, *J. Exp. Med.* **195**, 893 (2002).
24. S. A. Rosenberg et al., *N. Engl. J. Med.* **323**, 570 (1990).
25. M. Matloubian, R. J. Concepcion, R. Ahmed, *J. Virol.* **68**, 8056 (1994).
26. S. C. Jameson, *Nature Rev. Immunol.* **2**, 547 (2002).
27. R. Chiklis et al., *N. Engl. J. Med.* **343**, 750 (2000).
28. S. R. Riddell, P. D. Greenberg, *J. Immunol. Methods* **128**, 189 (1990).
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Materials and Methods

Fig. S1

Tables S1 and S2

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